NMDP CURRENT INVENTORY REQUIREMENTS FOR NEW CORD BLOOD UNITS Approved by the NMDP Cord Blood Committee May 3, 2004 Amended by the NMDP Cord Blood Committee September 27, 2004 Amended by the NMDP Cord Blood Committee January 27, 2006 Amended by the Cord Blood Committee October 5, 2007 Amended by the Cord Blood Committee Nov. 19, 2008 Amended by the Cord Blood Advisory Group Nov 2, 2012

This document is to be used in conjunction with requirements contained in the Cord Blood Bank Participation Agreement and NMDP Standards. All of the following requirements are intended for both NMDP member and listing cord blood banks prior to final individual unit registration into NMDP computer systems. (Please note specific member and listing requirements in items #1 and #2).

| Item | Item | Detail | Comment | | | |
|-----------------|--|---|--|--|--|--|
| Number | | | | | | |
| | Cectious Disease Marker Testing of Maternal Samples | | | | | |
| Infectious 1 | Disease Marker Testin Infectious Disease Marker (IDM) Testing | Member CBBsRequired tests:HBsAg, Anti-HCV, Anti-HIV 1/2 (or anti-HIV1/2 plus O), HIV NAT, anti-HBc, HCV NAT, Anti-HTLVI/I, Syphilis, anti-CMV Total,WNV NAT, and Chagas.HBV NAT testing is optional.Testing method:Testing must be performed usingFDA licensed, approved, orcleared donor screening testsperformed in a CMS approvedlaboratory in accordance withmanufacturer's instructions andin compliance with current tissueregulations.Listing CBBsAll applicable national and localregulations must be followed.Required tests:HBsAg, Anti-HCV, Anti-HIV 1/2 (or anti-HIV1/2 plus O), HIV NAT or p24.Where available, testing shouldbe performed for anti-HBc, HCVNAT, Anti-HTLV I/II, Syphilis,anti-CMV Total, WNV NAT,and Chagas. HBV NAT testingis optional.Testing method:It is preferred that the testing | Member and Listing CBBsTesting must be performed on a maternal sample collected within seven days (before or after) of the date of collection of the cord blood unit.All units must be screening test negative, with the exception of anti- HBc, Syphilis, and anti-CMV. Units that are anti-HBc positive will be accepted if HBsAg testing is negative. Units that are Syphilis screening test positive, confirmatory test (FTA- ABS) negative will be accepted.CMV test status does not impact | | | |
| | | method listed above for member CBBs is used if possible. | according to FDA criteria, the units will be managed as Exception or Incomplete depending on the date of collection of the cord blood unit. | | | |
| | | | | | | |

| Health History Screening | | | | | |
|--------------------------|----------------------------|--|---|--|--|
| 2 | Maternal and | Member CBBs | The NMDP MRQ and FMHQ are | | |
| | Family | Must have current donor | consistent with the FDA Guidance | | |
| | - | screening processes that are | for Industry Eligibility Determination | | |
| | | consistent with the FDA | for Donors of Human Cells, Tissues, | | |
| | | Guidance for Industry Eligibility | and Cellular and Tissue-Based | | |
| | | Determination for Donors of | Products (HCT/Ps) August 2007. | | |
| | | Human Cells, Tissues, and | CBB equivalent documents may also | | |
| | | Cellular and Tissue-Based | acceptable. AABB would also be | | |
| | | Products (HCT/Ps) August 2007. | considered as having an equivalent | | |
| | | Screening must address a current | MRQ. | | |
| | | maternal medical history, family | The intent of the screening process is | | |
| | | medical history, and other | to identify potential infectious | | |
| | | available medical records | disease risk, and inherited medical | | |
| | | including relevant and readily | conditions potentially transmitted | | |
| | | available physical findings and | though transplantation. | | |
| | | delivery records as defined by | Screening must be performed within | | |
| | | the cord blood bank. | 6 months prior to the time of | | |
| | | Listing CPPs | collection and must be performed in | | |
| | | Listing CBBs Must have current donor | a manner that provides current | | |
| | | screening processes that are | relevant history information. | | |
| | | consistent with applicable laws | | | |
| | | and regulations of the country of | | | |
| | | origin and address a current | | | |
| | | maternal medical history, family | | | |
| | | medical history, and other | | | |
| | | available medical records | | | |
| | | including relevant and readily | | | |
| | | available physical findings and | | | |
| | | delivery records as defined by | | | |
| | | the cord blood bank. | | | |
| | | llected at the Time of CBU Collecti | on | | |
| 3 | Serum or Plasma | Two vials of serum or non- | | | |
| | | heparinized plasma containing a | | | |
| | | minimum of 1.8 ml each should | | | |
| | | be stored at \leq -70°C. | | | |
| 4 | DNA | Material for preparation of ≥ 50 | The CBB may store frozen cellular | | |
| | | μ g of genomic DNA must be | material, extracted DNA or filter | | |
| | | collected and stored. | paper blots provided the material is | | |
| | | | equivalent to or sufficient to prepare $> 50 \text{ ug of genomic DNA}$ | | |
| Testing or | I I Final CBU Product J | Post-processing, Pre-freezing (prior | \geq 50 µg of genomic DNA. | | |
| 5 | Total Nucleated | TNC must be measured on a | Note: FDA licensure requires a | | |
| | Cell Count (TNC) | post-processing sample, reported | minimum post-processing TNC of 50 | | |
| | | as the number of cells $\times 10^7$ and | $\times 10^7$ | | |
| | | must <u>not</u> be corrected for | | | |
| | | nRBCs. | | | |
| 6 | Nucleated Red | nRBC must be determined on | The CBB must report the nRBC | | |
| | Blood Cell Count | the same sample as the post- | value as it is received from the | | |
| | (nRBC) | processing TNC and can be | testing laboratory. No additional | | |
| | | reported as either the % of total | calculations or conversions should be | | |
| | | or as an absolute count. | performed by the CBB. | | |
| • | • | • | · - · | | |

| 7 | CD34+ | The CD34+ count must be | Note: EDA licensure requires a |
|-----------|------------------------------|---|--|
| / | CD34+ | determined on the post- | Note: FDA licensure requires a minimum post-processing CD34+ |
| | | processing sample and reported | count of $\geq 1.25 \times 10^6$ viable CD34+ |
| | | as the total number of cells x | cells/unit. |
| | | 10 ⁶ . | |
| 8 | Colony Forming | CFU assays must be determined | CFU assays may include all colony |
| | Unit (CFU) Assay | on the post-processing sample | types or be limited to granulocyte- |
| | | and reported as "Growth" or "No | macrophage (GM). |
| | | Growth" with the actual count. | CBUs with no growth will not be |
| 9 | Viability | Viability must be determined on | listed. Viability may be performed by |
| 9 | v lability | the post-processing sample and | whatever assay is commonly used in |
| | | be \geq 85%. | the laboratory performing the test. |
| | | $bc \ge 0.5 / 0.$ | Note: FDA licensure requires $\geq 85\%$ |
| | | | viable nucleated cells (TNC). |
| Testing o | n CBU Residual Samp | les Post-processing | |
| 10 | Bacterial and | CBUs must be tested for | Testing may be performed on a |
| | Fungal Culture | aerobic/anaerobic microbes and | residual red blood cell and/or plasma |
| | | fungus and must exhibit "No | sample remaining after processing, |
| | | Growth". | on the final product prior to |
| | | | cryopreservation, or on the final |
| | | | product after addition of DMSO. |
| | | | When the culture is performed before |
| | | | the addition of DMSO, evidence that |
| | | | the DMSO was sterile must be |
| 11 | I I ann a al a h-in an athai | Testing for home slabin mothing | documented. |
| 11 | Hemoglobinopathy | Testing for hemoglobinopathies, including sickle cell disease and | Testing may either be performed on residual rbc material remaining post- |
| | Testing | thalassemia, must be performed | processing or on a sample of whole |
| | | prior to listing CBUs. The test | cord blood prior to processing. |
| | | must utilize a method that | Newborn screening is also acceptable |
| | | distinguishes sickle cell disease | provided that the results are |
| | | and trait, alpha and beta | available. A solubility assay for |
| | | thalassemia disease and trait and | hemoglobin screening is not |
| | | Hemoglobin C disease. CBUs | acceptable. |
| | | homozygous for either sickle | - |
| | | cell disease or thalassemia will | CBUs testing positive for multiple |
| | | be deferred. CBUs heterozygous | traits may be deferred. However, a |
| | | for either sickle cell trait or | particular unit with multiple traits |
| | | thalassemia will be accepted. | may be used upon consultation with |
| | | | an expert in the field of |
| | | | hemoglobinopathies and with the |
| | | | knowledge and approval of the |
| 12 | HLA Typing | CBUs must be typed using DNA | transplant center. HLA typing may either be performed |
| 14 | TILA Typing | methodology at a minimum of | on residual material containing |
| | | low resolution for HLA-A and - | nucleated cells following processing |
| | | B, and high resolution for HLA - | or on a sample of whole cord blood |
| | | DRB1. | prior to processing. |
| | | | |
| 13 | ABO Rh Typing | CBUs must be typed for ABO | ABO/Rh typing may either be |
| | | and Rh. | performed on residual rbc material |
| | | | remaining post processing or on a |
| | | | sample of whole cord blood prior to |
| | | | processing. |
| | 1 | | |

| Storage of CBU | | | | | |
|----------------|--|---|--|--|--|
| 14 | Temperature | CBUs must be stored at temperatures $\leq -150^{\circ}$ C at all times. | Storage may occur in the liquid or vapor phase of nitrogen. | | |
| Storage of | f Cord Blood Samples | | | | |
| 15 | Serum/Plasma | Two vials of serum or non- heparinized plasma containing a minimum of 1.8 ml each should be stored at \leq -70°C. | The material for these samples may come from residual material (undiluted) following volume reduction. | | |
| 16 | DNA | Material for preparation of ≥ 50 µg of genomic DNA must be collected and stored. | The CBB may store frozen cellular material, extracted DNA or filter paper blots provided the material is equivalent to or sufficient to prepare $\geq 50 \ \mu g$ of genomic DNA. | | |
| Viable Ce | ell Requirements (CBU | | | | |
| 17 | Viable Cells Aliquot = sample representative of the final product | A minimum of three aliquots of cells each containing a minimum of 1×10^6 cells must be cryopreserved and stored at \leq - 150° C. | Aliquots must include at least two contiguous segments – additional aliquots may include any that contain a cryoprotectant and are stored in the same manner as the CBU. Note: For CBBs in the US, licensure requirements may indicate that an appropriately labeled reserve sample representative of the final product is maintained as a reserve/retention sample, beyond the minimum requirements specified here, consistent with a CBB's Biologic License Application (BLA). | | |
| 18 | Contiguous Segments | All CBUs must have at least two contiguous segments containing a minimum of 100 µl each. | One segment will be used for HLA confirmatory typing. One segment (additional segment or same as HLA segment if enough material) will be used for additional testing at the time of release of the unit for transplant (ex. TNC, viability, CFU). Alternatively, TNC and viability may be performed from an aliquot stored separately from the CBU. Every attempt should be made to leave one segment attached to the unit for shipment to the transplant center. Note: For CBBs in the US, licensure requirements may indicate that an appropriately labeled reserve sample representative of the final product is maintained as a reserve/retention sample, beyond the minimum requirements specified here, consistent with a CBB's Biologic License Application (BLA). | | |